



## INVESTIGATION OF *Melaleuca leucadendron* L. FROM SOUTH SUMATRA: Phytochemicals and IC50-BASED ANTIOXIDANT POTENTIAL

Rima Ernia<sup>1</sup>, Rina SE Sitindaon<sup>1</sup>, Nyoman Yudi Antara<sup>2\*</sup>

<sup>1</sup> Medical Laboratory Technology Study Program, Faculty of Health, Kader Bangsa University, Palembang, Indonesia

<sup>2</sup> Biotechnology Study Program, Faculty of Mathematics and Sciences, Universitas Halu Oleo, Kendari Indonesia

\* Corresponding author : [yudiantaranyoman@uho.ac.id](mailto:yudiantaranyoman@uho.ac.id)

Received : Oktober 2025

Revised : November 2025

Accepted : Desember 2025

First Publish Online :  
Desember, 30, 2025

Keywords : *Melaleuca leucadendron L*,  
Phytochemical Screening,  
Antioxidant Activity

### ABSTRACT

*Melaleuca leucadendron L.* (commonly known as Gelam wood) is a plant that thrives in the swampy regions of South Sumatra. The utilization of this plant has historically been limited to its wood, leaving the potential of its leaves and seeds largely unexplored. Therefore, this study aimed to identify the species of Gelam wood and investigate the phytochemical content and antioxidant potential of its leaves and seeds. The methodology involved methanol extraction of the plant material, followed by comprehensive phytochemical screening and an antioxidant activity assay using the DPPH method. The phytochemical analysis confirmed the presence of several key compounds, with the Gelam extracts testing positive for Alkaloids, Flavonoids, Steroids, Saponins, Tannins, and Triterpenoids. Furthermore, the extract from the leaves and seeds demonstrated potent antioxidant activity, evidenced by an IC50 value of 47.56 ppm, which classifies it as a strong antioxidant (<50 ppm). This research highlights the significant therapeutic potential of *Melaleuca leucadendron* leaves and seeds, suggesting they are valuable sources of natural antioxidants.

This is an open-access article under the [CC-BY-SA](#) license



## Introduction

Exploration into the potential of plant resources within Indonesia continues actively to this day. This potential is strongly supported by the country's rich biodiversity, which means that every plant species in a given region possesses unique characteristics and harbors many undiscovered benefits (Setiawan, 2022). Nowadays, natural antioxidants of plant is a focus as alternatives to synthetic counterparts that is for improving oxidation stability (Lankanaya A, 2025). Often, these

valuable plants grow right in our surroundings without their potential being fully realized. One such plant commonly used by the people of South Sumatra primarily for its timber, yet whose full benefits are unknown, is Gelam wood (*Melaleuca leucadendron*). Gelam wood is easily found in swampy regions (Basyaruddin et al., 2019; Istanto & Saputra, 2013). Its presence is significant in South Sumatra due to the ideal habitat for its growth. Currently, Gelam wood in South Sumatra is exploited mainly for its high-

quality timber, while its leaves and seeds have not been widely utilized (Pramono, 2016). Therefore, successfully utilizing the leaves and seeds of Gelam wood would enhance the plant's economic value and foster new economic activities for local communities.

Gelam wood is recognized as belonging to the same family as Cajeput wood (*Melaleuca cajuputi*), a plant famously utilized in the production of essential oils (Aryani, 2020). While the essential oil derived from the cajeput tree has been well-known and used by communities for a long time, the potential of the closely related Gelam tree has not yet been fully explored. To address this gap and determine the potential of Gelam wood, particularly in its leaves and seeds, this research was specifically conducted to investigate the phytochemical content and antioxidant capacity of methanol extracts derived from these parts of the Gelam plant.

## Materian and Methods

### Extraction Method

The extraction was performed using the maceration method. A 500 gram sample of the dried powder was submerged in ethanol at a 1:1 ratio (powder:solvent). This mixture was allowed to stand for 72 hours in a cool place, protected from direct light, with stirring performed every 3 hours for 5 minutes throughout the period. After 72 hours, the resulting macerate was filtered using a funnel lined with Whatman No. 1 filter paper. The filtrate was collected and subjected to rotary evaporation at a temperature of 55 °C to separate the solvent from the crude extract. Finally, the remaining extract was concentrated further using a water bath to yield a viscous extract (Hakim et al., 2019).

### Phytochemical Screening of Leaf and Seed Simplicia of *Melaleuca leucadendron*

Tannin testing was initiated by dissolving 5 g of the dried leaf and seed material (simplicia) in a beaker containing 20 mL of

distilled water (a *qua des*), followed by five minutes of heating and subsequent filtration. The resulting filtrate was then treated with three drops of 10% Iron(III) Chloride ( $FeCl_3$ ) solution, with a positive result indicated by the formation of a blackish-green color. The Saponin test used the same initial preparation; however, the filtrate was transferred to a tightly capped test tube and shaken vigorously. The test was considered positive if a stable foam (emulsion) was produced.

For the Flavonoid test, 5 g of the dried leaf and seed material (simplicia) was dissolved in a beaker containing 20 mL of distilled water (a *qua des*). After heating for five minutes, the resulting filtrate was transferred to a test tube, followed by the addition of Magnesium powder, a 1:1 mixture of HCl (hydrochloric acid) and EtOH (ethanol), and finally, amyl alcohol. A positive result was indicated if an orange layer formed in the amyl alcohol phase. Conversely, for the Steroid test, 1 g of the sample was extracted with hot EtOH and filtered. The filtrate was heated to dryness, and the residue was treated with 1 mL of diethyl ether, one drop of concentrated  $H_2SO_4$  (sulfuric acid), and one drop of concentrated anhydrous  $CH_3COOH$  (acetic acid). The test was considered positive upon the formation of a green or blue color. For the Triterpenoid test, 1 g of the sample was placed in a test tube, extracted with hot EtOH (ethanol), and filtered. The filtrate was evaporated to dryness, and the residue was then treated with 1 mL of diethyl ether. This solution was subsequently treated with one drop of  $H_2SO_4$  (sulfuric acid) and one drop of concentrated  $CH_3COOH$  (acetic acid). A positive result was indicated by the formation of a red or purple color. Separately, the Alkaloid test was initiated by grinding 1 g of the sample in a mortar with a few drops of  $NH_3$  (ammonia). The resulting paste was extracted with 5 mL of  $CHCl_3$  (trichloromethane) and filtered. The filtrate was mixed with 2 M  $H_2SO_4$  and shaken regularly, then allowed to settle

until three distinct layers formed. The test was considered positive if the following precipitates were observed in the first layer: a reddish-brown precipitate with Wagner's reagent.

### Antioxidant Activity Assay

The assay began with the preparation of a 40 ppm DPPH solution by dissolving 0.01 g of DPPH in a 250 mL volumetric flask using methanol, which was then kept cold and protected from light. The solution's maximum wavelength ( $\lambda_{\text{max}}$ ) was determined across 490–534 nm using methanol as a blank. For the radical scavenging activity determination, 2 mg of the methanol extract was used to prepare test solutions at six concentrations ranging from 3.125 to 100 ppm. A quercetin standard was prepared at the same concentrations. 1 mL of each test solution was mixed with 2 mL of the 40 ppm DPPH solution and incubated for 30 minutes at room temperature. Absorbance was subsequently measured at 517 nm using a UV-Vis spectrophotometer. The experiment included two measurements and three replicates for each sample, after which the IC<sub>50</sub> value—the concentration

Table 1. Phytochemical parameters for positive result

| Ingredients of Phytochemical Compounds | Interpretation   | Reference  |
|--|--|--|
| Alkaloids                              | Formed a brown precipitate   | Formed a brown precipitate with Dragendorff reagent (Kopon et al., 2020)                       |
| Flavonoids                             | A red color is formed on the layer   | Red, yellow or orange color is formed (Kopon et al., 2020)                                     |
| Saponins                               | Formed froth that persisted even though HCl 2 N was added (foam did not disappear) | Stable froth formed even though 2N HCl was added (foam did not disappear) (Kopon et al., 2020) |
| Tannins                                | Formed a black green color   | Formed a black-green color (Kopon et al., 2020)  |
| Steroids                               | Blackish green formed  | Blackish green color is formed (Putra et al., 2016)  |
| Triterpenoids                          | Blackish purple formed   | Black purple color is formed (Putra et al., 2016)  |

Antioxidant capacity is quantified by measuring the decrease in absorbance of the DPPH solution after the sample is added. This change is calculated as the percentage of inhibition (% Inhibition) by comparing

required to inhibit 50% of DPPH activity—was calculated using the regression equation, where a smaller IC<sub>50</sub> value indicates higher antioxidant activity (Salazar-Aranda et al., 2011).

### Research Location and Time

The determination test was conducted at the Biology Laboratory, Andalas University, while phytochemical testing was carried out at the Pharmacy Laboratory, Kader Bangsa University. Subsequently, the antioxidant activity analysis was performed at the Biology Laboratory, Sriwijaya University. The sample was collected from swamp located within Banyuasin regency. This research was completed in 2022.

### Experimental Design

The research design employed an experimental design, and the sampling method used was simple random sampling.

### Research Methods and Parameters

The parameter for phytochemicals testing following the previous method and described in table 1.

the absorbance before and after adding the extract. The resulting inhibition percentages are then plotted against the corresponding extract concentrations (ppm) to generate a linear regression equation in

the form of  $Y=a+bX$ , where  $X$  is the concentration (X-axis) and  $Y$  is the percentage of inhibition (Y-axis). The crucial IC<sub>50</sub> value is determined from this equation by calculating the extract concentration (X) required to achieve a 50% inhibition (Y=50), serving as the

standard metric for expressing antioxidant strength (Salazar-Aranda et al., 2011).

### Data Analysis

The phytochemical analysis was interpreted qualitatively, whereas antioxidant activity results were subjected to linear regression analysis to determine the IC<sub>50</sub> value.

## Results and Discussion

### Results

The leaves of Gelam wood (*Melaleuca leucadendra* (L.) L.) are typically oblong or lanceolate (4.5 cm to 15 cm long), possess a hairy texture ranging in color from green

to brownish, and emit a distinct cajeput oil fragrance when crushed. The plant produces compound spike inflorescences with bell-shaped, white flowers that emerge at the branch tips, leading to small, light-to-dark brown fruit (2.5 mm to 4 mm wide, as shown in Figure 1)

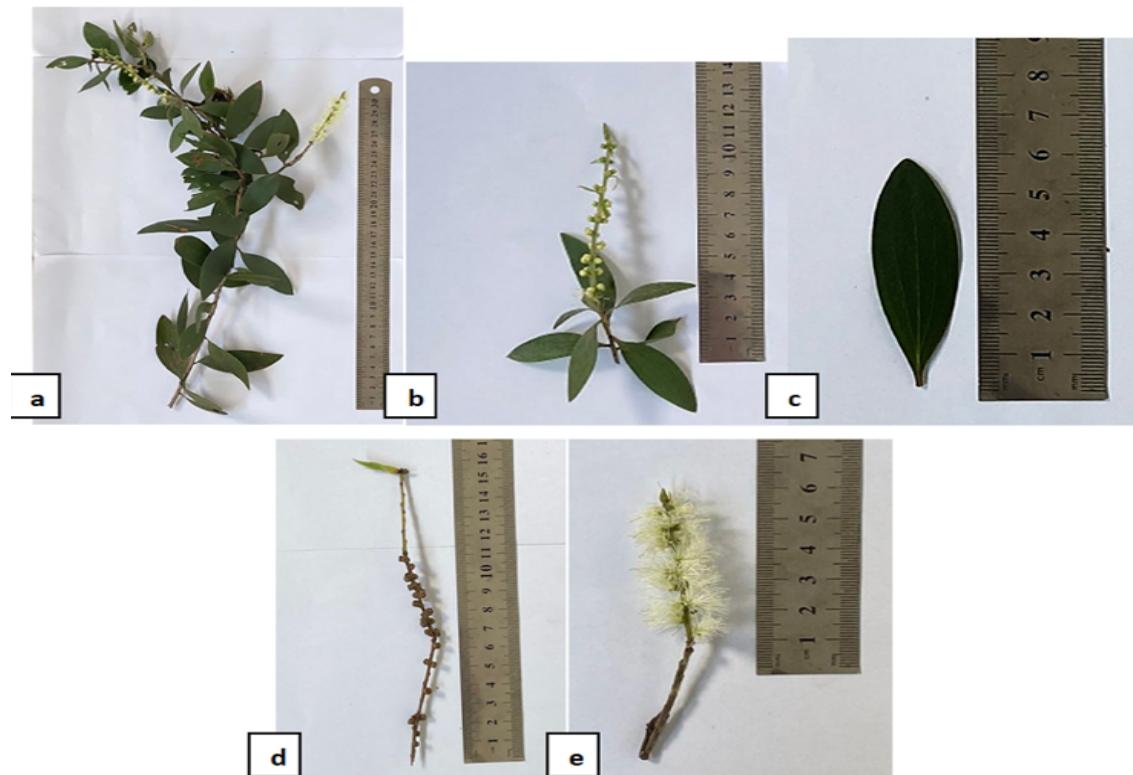


Figure 1. Morphology of Gelam Wood (*Melaleuca leucadendra* (L.) a) Leaf Petiole and Inflorescence (Flower Spike) (b) Immature Fruit/Seed Capsule (c) Leaf(d) Mature Fruit/Seed Capsule (e) Flower

The identification of Gelam wood's content began with extraction. The leaves and seeds were extracted using the maceration method with methanol as the solvent. The

maceration was performed at room temperature in the dark for a total period of 3×24 hours (72 hours). The resulting crude extract from the leaves and seeds was a

viscous material with a blackish-green color and a distinct cajeput oil-like fragrance. From an initial sample of 400 g

of dried leaf and seed powder macerated in 12 L of methanol, the final evaporation yielded 154 g of concentrated extract.

Table 2. Results of Phytochemical Compound Identification in Gelam Wood Leaf and Seed Extracts

| Bioactive compound | Reagent                    | Inference |
|--------------------|----------------------------|-----------|
| Alkaloid           | Wagner                     | +         |
| Flavonoid          | Shinoda's test             | +         |
| Saponin            | Foam test                  | +         |
| Tanin              | FeCl3                      | +         |
| Steroid            | Liebermann-Burchard's test | +         |
| Triterpenoid       | Liebermann-Burchard's test | +         |



Figure 2: Phytochemical Screening Results. Showing positive identification for Alkaloids (A), Flavonoids (B), Saponins (C), Tannins (D), Steroids (E), and Triterpenoids (F).

The antioxidant activity of the Gelam wood leaf and seed extracts was measured using the widely favored DPPH (2,2-diphenyl-1-picrylhydrazyl) method, which is simple, fast, and requires minimal sample (Safitri et al., 2020). The method's principle is based on the DPPH radical's ability to accept a hydrogen atom donated by an antioxidant compound. This reaction is monitored

spectrophotometrically at 517 nm (the maximum absorption wavelength of DPPH) by observing the decrease in absorbance as the solution changes color from violet to the more stable, yellow DPPH-Hydrazine form; consequently, the intensity of the color change is directly proportional to the extract's antioxidant capacity (Molyneux, 2003).

Table 3. Percentage Inhibition of Quercetin

| Concentration (µg/mL) | Ln Concentration | % Inhibition |        |        |
|-----------------------|------------------|--------------|--------|--------|
|                       |                  | 1            | 2      | 3      |
| A 100                 | 4.6052           | 86.269       | 86.008 | 86.269 |
| B 50                  | 3.9120           | 85.601       | 85.630 | 85.630 |
| C 25                  | 3.2189           | 85.281       | 85.281 | 85.223 |
| D 12.5                | 2.5257           | 84.176       | 84.496 | 84.002 |
| E 6.25                | 1.8326           | 82.461       | 82.955 | 83.014 |
| F 3.125               | 1.1394           | 82.112       | 81.415 | 82.171 |

Quercetin

33

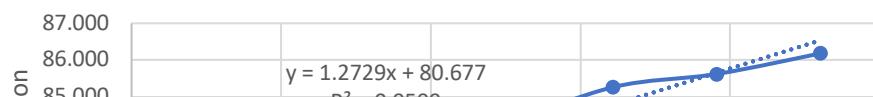


Figure 3: Calibration Curve Showing DPPH Absorbance vs. Quercetin Concentration  
The IC50 calculation results are exponential -24.10009 is 3.42 ppm

Tabel 4. Percentage of inhibition of antioxidant activity of Gelam Wood (leaves-seeds extract)

| Concentration<br>( $\mu\text{g/mL}$ ) | Ln Concentration | % Inhibition |        |        |
|---------------------------------------|------------------|--------------|--------|--------|
|                                       |                  | 1            | 2      | 3      |
| A 1000                                | 6.9078           | 87.694       | 75.581 | 78.779 |
| B 500                                 | 6.2146           | 78.682       | 61.047 | 65.698 |
| C 250                                 | 5.5215           | 65.310       | 55.814 | 57.849 |
| D 125                                 | 4.8283           | 57.461       | 47.384 | 44.186 |
| E 62.5                                | 4.1352           | 54.554       | 40.116 | 35.756 |
| F 31.25                               | 3.4420           | 48.740       | 37.791 | 37.151 |

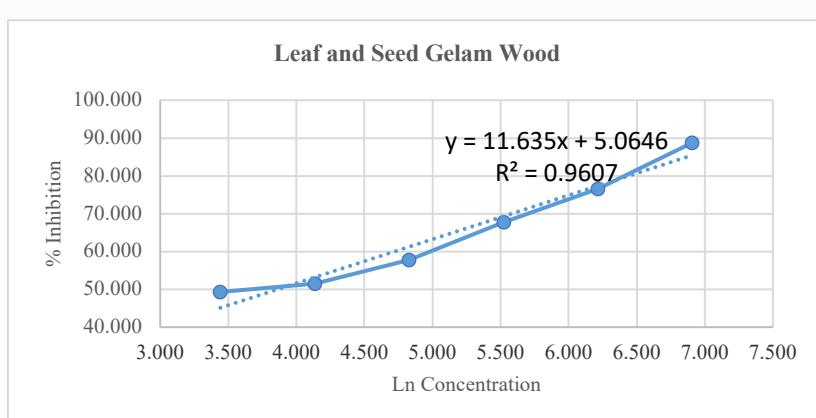


Figure 4: DPPH Absorbance vs. Concentration Graph for Gelam Wood Leaf and Seed Extracts  
The IC50 calculation results are as x = 3.862 exponential 3.862 is 47.565 ppm

## Discussion

This resilient species thrives extensively in peat swamp forests, particularly well in eastern Indonesia and northern Australia, though it is also cultivated elsewhere with clear dry seasons. It is commonly found in barren soils with a

pH of 5.6, is heat-tolerant, and can resprout after fire, growing from sea level up to 400 meters above sea level near beaches, behind mangrove forests, or in various wet or dry swampy lands (Supriyati et al., 2014). The comprehensive results of the phytochemical identification are presented

in Table 2. The findings from the preliminary tests demonstrate that the methanol extract of the Gelam wood leaves and seeds contains alkaloid compounds. Alkaloids are significant secondary metabolites widely distributed across nature, found not only in plants but also in fungi and animals (Gwaltney-Brant et al., 2012; Maisarah et al., 2023). In plants, these compounds primarily function as toxic agents to deter insects that might otherwise harm the organism (Matsuura & Fett-Neto, 2015). In the field of pharmacology, alkaloids are highly valued for their therapeutic properties, serving as potent agents for anesthesia, cardioprotection, and anti-inflammation (Kurek, 2019). Furthermore, alkaloids are frequently utilized in pharmaceutical applications, including the development of various medicines and even some toxins (Roy, 2017).

The flavonoid phytochemical test on the Gelam wood leaf and seed extract yielded a positive result, evidenced by the formation of a red color. Flavonoids, known for their antibiotic properties, are functional in treating liver disorders and can act as hepatoprotectors by inhibiting prostaglandin synthesis (Dwika et al., 2016). Similarly, the phytochemical test for saponins was also positive, demonstrated by the presence of stable foam during the experiment. Saponins are essential antimicrobial agents whose concentration in plants is affected by variety and growth stage, with the highest levels typically found in the parenchymal cells of vegetative parts (Faizal & Geelen, 2013). Saponins promote regeneration and re-epithelialization by supporting the host's immune response. In pharmaceutical applications, they are valued as anticarcinogenic and immunostimulant agents, with further uses spanning agriculture, cosmetics, and food production (Moghimipour & Handali, 2015).

The tannin phytochemical test yielded a positive result, evidenced by the formation of a blackish-green color resulting from the reaction between the phenolic hydroxyl groups in tannins and  $\text{FeCl}_3$  (Iron(III) Chloride) to form an  $\text{Fe}^{3+}$  complex; tannins are recognized as pharmacologically active components (Oyetayo, 2007). The steroid phytochemical test was also positive, showing a green color that confirms the presence of steroid compounds; these compounds are vital for controlling plant growth and are used in medicine as detoxifying agents (Dwika et al., 2016). Finally, the triterpenoid phytochemical test was confirmed by a color change to purple-black, indicating the presence of triterpenoids, which primarily function in the plant as a defense mechanism against disruptive insects.

Antioxidant activity in this study is quantified by the  $\text{IC}_{50}$  value, which represents the concentration of the test compound required to scavenge 50% of free radicals; a smaller  $\text{IC}_{50}$  value indicates higher antiradical activity (Das, 2020). The  $\text{IC}_{50}$  for the Gelam wood leaf and seed extract was determined from the linear regression equation shown in Figure 4:  $y=11.635x+5.0646$  ( $r=0.9607$ ). By substituting 50 for the  $y$  variable, the concentration ( $x$ ) was calculated to be 47.56 ppm, meaning the extract scavenges 50% of free radicals at this concentration. Although this value is higher than the quercetin control (3.42 ppm), its  $\text{IC}_{50}$  remains below 50 ppm, which is the recognized threshold for classifying a substance as a strong antioxidant, thereby confirming the potent antioxidant content of the Gelam wood leaf and seed extracts. The antioxidant capacity of natural materials can vary significantly due to several factors, including the structure of the polyphenols present and the inherent, often weak, stability of those compounds. This stability is highly susceptible to chemical and physical factors such as pH, temperature, light, oxygen, enzymes,

ascorbic acid, sugars, and metal ions (Lang et al., 2024). Additionally, research on plants like *Hyssopus officinalis*, *Lamium album*, and *Leonurus cardiaca* has shown that the highest antioxidant content is often concentrated in the upper parts of the plant or tissues close to the flowers (Skrovankova & Mlcek, 2025). Consistent with these findings, the strong antioxidant content identified in this study was derived from the Gelam wood leaves and seeds, without dismissing the vital role that chemical and physical factors play in maintaining this antioxidant potency.

## Conclusions

Based on the determination test, the species of the Gelam wood plant was confirmed as *Melaleuca leucadendra*. The subsequent phytochemical screening revealed that the plant contains a diverse profile of bioactive compounds, including alkaloids, flavonoids, saponins, tannins, steroids, and triterpenoids. Furthermore, the leaf and seed extracts demonstrated strong antioxidant activity, evidenced by an IC<sub>50</sub> value of 47.56 ppm, confirming their ability to scavenge 50% of free radicals at this concentration. This study contributes new knowledge regarding the potential of Gelam wood. Future research should focus on a more in-depth quantitative analysis of its phytochemical content and further investigate its potential as an antifungal or antibacterial agent.

## Acknowledgment

The author extends sincere gratitude to all contributors to this study. Furthermore, this research was financially supported by a grant from RISTEKDIKTI.

## References

Aryani, F. (2020). Pengenalan Atsiri (Melaleuca cajuputi) Cara Poduksi dan Pengujian. *Jurusan Teknologi Pertanian Politeknik Pertanian Negeri Samarinda*, 1–38.

Basyaruddin, Suryaningsih, A., & Awali, J. (2019). *Potensi Pemanfaatan Kayu Gelam Dan Kayu Sengon Dalam Dunia Konstruksi Berdasarkan Uji Kuat Lentur*. 13(3), 193–198.

Das, U. N. (2020). *Introduction to Free Radicals, Antioxidants, Lipid Peroxidation, and Their Effects on Cell Proliferation BT - Molecular Biochemical Aspects of Cancer* (U. N. Das (ed.); pp. 41–65). Springer US. [https://doi.org/10.1007/978-1-0716-0741-1\\_2](https://doi.org/10.1007/978-1-0716-0741-1_2)

Dwika, W., Putra, P., Agung, A., Oka Dharmayudha, G., & Sudimartini, L. M. (2016). Identifikasi Senyawa Kimia Ekstrak Etanol Daun Kelor (*Moringa oleifera* L) di Bali (Identification Of Chemical Compounds Ethanol Extract Leaf Moringa (*Moringa Oleifera* L) In Bali). *Indonesia Medicus Veterinus Oktober*, 5(5), 464–473.

Faizal, A., & Geelen, D. (2013). Saponins and their role in biological processes in plants. *Phytochemistry Reviews*, 12(4), 877–893. <https://doi.org/10.1007/s11101-013-9322-4>

Gwaltney-Brant, S. M., Dunayer, E., & Youssef, H. (2012). *Chapter 73 - Terrestrial zootoxins* (R. C. B. T.-V. T. (Second E. Gupta (ed.); pp. 969–992). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-385926-6.00084-3>

Hakim, R. I., Wilson, W., & Darmawati, S. (2019). Uji Aktivitas Antibakteri Ekstrak Ethanol Daun Kayu Putih (*Melaleuca leucadendron* L.) terhadap Pertumbuhan Methicillin Resistant *Staphylococcus aureus* (MRSA) Antibacterial Test Activities of White Wood Ethanol Extract (*Melaleuca leucadendron* L.) on Growth. *Prosiding Seminar Nasional Mahasiswa Unimus*, 2, 109–115. <http://prosiding.unimus.ac.id>

Istanto, K., & Saputra, A. E. (2013). Analisis Model Pondasi Bangunan Air

di Atas Tanah Rawa Berbahan Lokal (Analysis of Waterworks Foundation Model on Swamp Soil Using Local Material). *TekTan Jurnal Ilmiah Teknik Pertanian*, 5, 135–202.

Kopon, A. M., Baunsele, A. B., & Boelan, E. G. (2020). Skrining Senyawa Metabolit Sekunder Ekstrak Metanol Biji Alpukat (*Persea Americana* Mill.) Asal Pulau Timor. *Akta Kimia Indonesia*, 5(1), 43. <https://doi.org/10.12962/j25493736.v5i1.6709>

Kurek, J. (2019). *Alkaloids: their importance in nature and human life*. BoD–Books on Demand.

Lang, Y., Gao, N., Zang, Z., Meng, X., Lin, Y., Yang, S., Yang, Y., Jin, Z., & Li, B. (2024). Classification and antioxidant assays of polyphenols: a review. *Journal of Future Foods*, 4(3), 193–204. <https://doi.org/10.1016/j.jfutfo.2023.07.002>

Lankanaya, A., Lakshan, N.D., Jayathunge, L., Bandara, P., Manatunga, D.C., Senanayake, C.M. (2025). A review of sustainable strategies for encapsulating antioxidant-rich plant polyphenolic extracts using nanoemulsification to enhance the oxidative stability of edible oils: *Discover Food*, 5(65), <https://doi.org/10.1007/s44187-025-00331-8>

Maisarah, M., Chatri, M., & Advinda, L. (2023). Karakteristik dan Fungsi Senyawa Alkaloid sebagai Antifungi pada Tumbuhan. *Jurnal Serambi Biologi*, 8(2), 231–236.

Matsuura, H. N., & Fett-Neto, A. G. (2015). *Plant Alkaloids: Main Features, Toxicity, and Mechanisms of Action BT - Plant Toxins* (P. Gopalakrishnakone, C. R. Carlini, & R. Ligabue-Braun (eds.); pp. 1–15). Springer Netherlands. [https://doi.org/10.1007/978-94-007-6728-7\\_2-1](https://doi.org/10.1007/978-94-007-6728-7_2-1)

Moghimpour, E., & Handali, S. (2015). Saponin: Properties, Methods of Evaluation and Applications. *Annual Research & Review in Biology*, 5(3), 207–220. <https://doi.org/10.9734/arrb/2015/11674>

Molyneux, P. (2003). The use of the stable free radical diphenylpicryl- hydrazyl ( DPPH ) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*, 50(June 2003).

Oyetayo, V. O. (2007). Comparative Studies of Phytochemical and Antimicrobial Properties of the leaf, stem and Tuber of Anchomanes difformis. In *journal of Pharmacology and Toxicology* (Vol. 2, Issue 4, pp. 407–410).

Pramono, A. A. (2016). Variasi produksi benih gelam (*Melaleuca leucadendron*) pada beberapa tegakan di Sumatera Selatan. December 2016. <https://doi.org/10.13057/psnmbi/m020204>

Putra, I. P., Dharmayudha, A. A. G. O., & Sudimartini, L. M. (2016). Identifikasi Senyawa Kimia Ekstrak Etanol Daun Kelor (*Moringa oleifera* L) di Bali. *Indonesia Medicus Veterinus*, 5. <https://api.semanticscholar.org/CorpusID:89718712>

Roy, A. (2017). A Review On The Alkaloids an Important Therapeutic Compound From Plants. *International Journal of Plant Biotechnology*, 3(2), 1–9. [https://www.researchgate.net/publication/320098967\\_A\\_review\\_on\\_the\\_alkaloids\\_an\\_important\\_therapeutic\\_compound\\_from\\_plants%0Ahttps://www.researchgate.net/profile/Arpita-Roy-3/publication/320098967\\_A\\_review\\_on\\_the\\_alkaloids\\_an\\_important\\_therapeutic\\_compoun](https://www.researchgate.net/publication/320098967_A_review_on_the_alkaloids_an_important_therapeutic_compound_from_plants%0Ahttps://www.researchgate.net/profile/Arpita-Roy-3/publication/320098967_A_review_on_the_alkaloids_an_important_therapeutic_compoun)

Safitri, F. W., Abdul, A., & Qonitah, F. (2020). Antioxidant activity test of

fennel leaves ethanol extract (*Foeniculum vulgare* Mill) using DPPH and FRAP methods. *Journal of Pharmaceutical Science and Medical Research*, 3(2), 43–54. <http://e-journal.unipma.ac.id/index.php/pharmmed>

Salazar-Aranda, R., Pérez-López, L. A., López-Arroyo, J., Alanís-Garza, B. A., & Waksman De Torres, N. (2011). Antimicrobial and antioxidant activities of plants from northeast of Mexico. *Evidence-Based Complementary and Alternative Medicine*, 2011. <https://doi.org/10.1093/ecam/nep127>

Setiawan, A. (2022). Keanekaragaman Hayati Indonesia: Masalah dan Upaya Konservasinya. *Indonesian Journal of Conservation*, 11(1), 13–21. <https://doi.org/10.15294/ijc.v11i1.34532>

Skrovankova, S., & Mlcek, J. (2025). Antioxidant Potential and Its Changes Caused by Various Factors in Lesser-Known Medicinal and Aromatic Plants. *Horticulturae*, 11(1). <https://doi.org/10.3390/horticulturae11010104>

Supriyati, W., Prayitno, T. A., Marsoem, S. N., Kehutanan, J., Pertanian, F., Palangka Raya, U., Raya, P., Kehutanan Universitas Gadjah Mada, F., & Yogyakarta, B. (2014). Proporsi Kayu Teras dan Sifat Fisik-Mekanik pada Tiga Kelas Diameter Kayu Gelam (*Melaleuca* sp) dari Kalimantan Tengah (Heartwood Proportion and Physical-Mechanical Properties of Three Diameter Class of Gelam Wood (*Melaleuca* sp) from Central Kalimantan). *J. Ilmu Dan Teknologi Kayu Tropis*, 12(1), 56–64. <https://jurnal.ugm.ac.id/JML/article/view/18729/12022>